

An Adaptive Framework for Non-target Risk Assessment of RNAi-based, Insect Resistant GM Crops

Jonathan Lundgren
Ecdysis Foundation
Estelline, SD



Dr. Kelton Welch



Dr. Chrissy Mogren

Environmental Risk Assessment

Proving a negative

No amount of
experimentation can
ever prove me right; a
single experiment can
prove me wrong.



Albert Einstein
German Theoretical-Physicist
(1879-1955)

QuoteHD.com

*“GM crops do not pose an extraordinary risk to
the environment”*

RNAi

*RNAi is a post-transcriptional technique for **sequence-selective** silencing of genes*

A concern is unintended gene
silencing in non-target
organisms

Lundgren and Duan. 2013. Bioscience 63: 657

For an animation on RNAi and associated risks,
search youtube for “Cable Hardin RNAi”



Cable Hardin, SD State

How Do We Select Species for Risk Assessment?



You can't measure all of the species in a habitat.



Indicator species

- Represent a functional or phylogenetic "guild"
- Are easy to rear and assay

Bioinventories and Risk Assessment

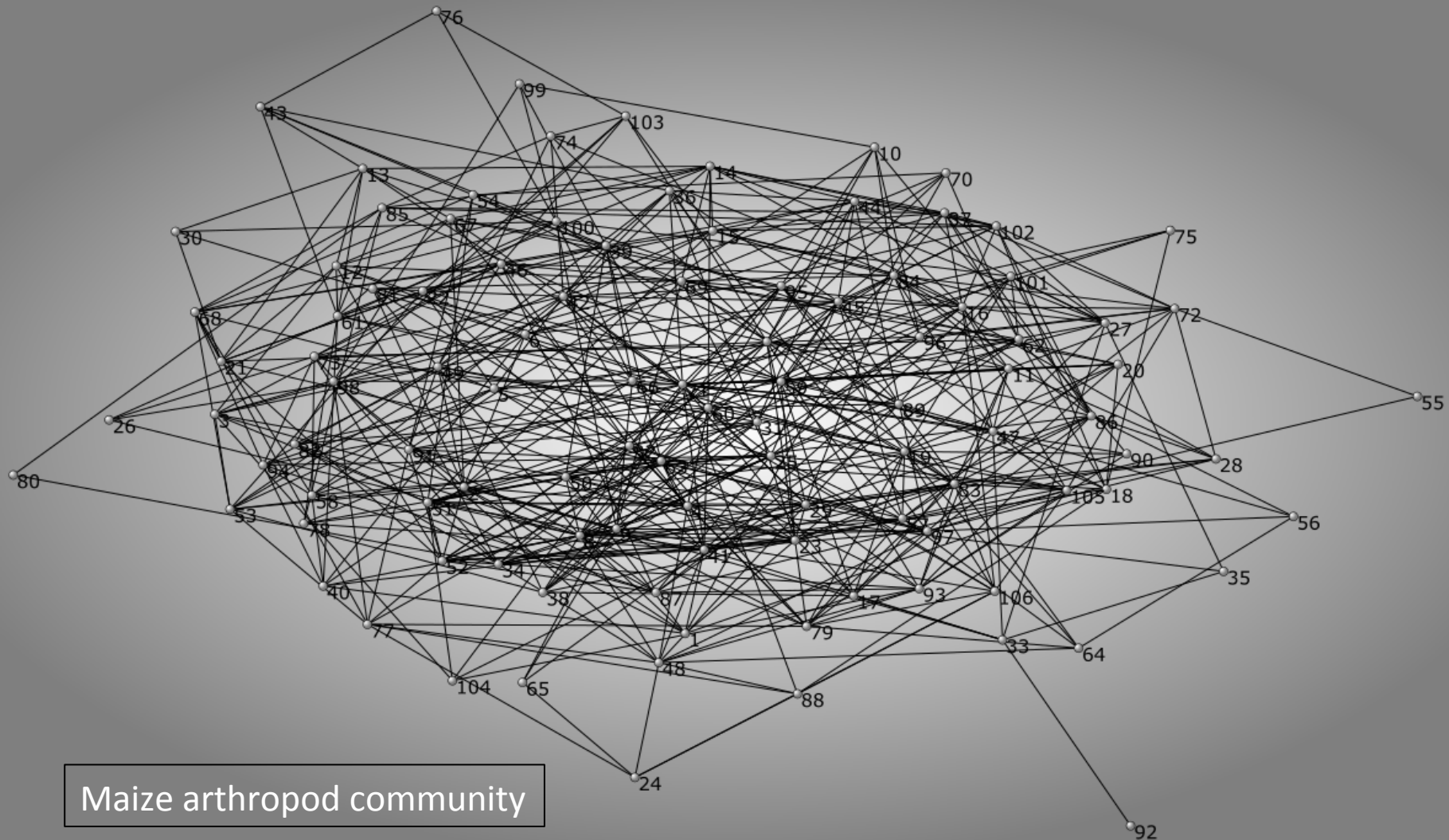
We have a poor understanding of the species that occur in most agroecosystems

Taxonomy

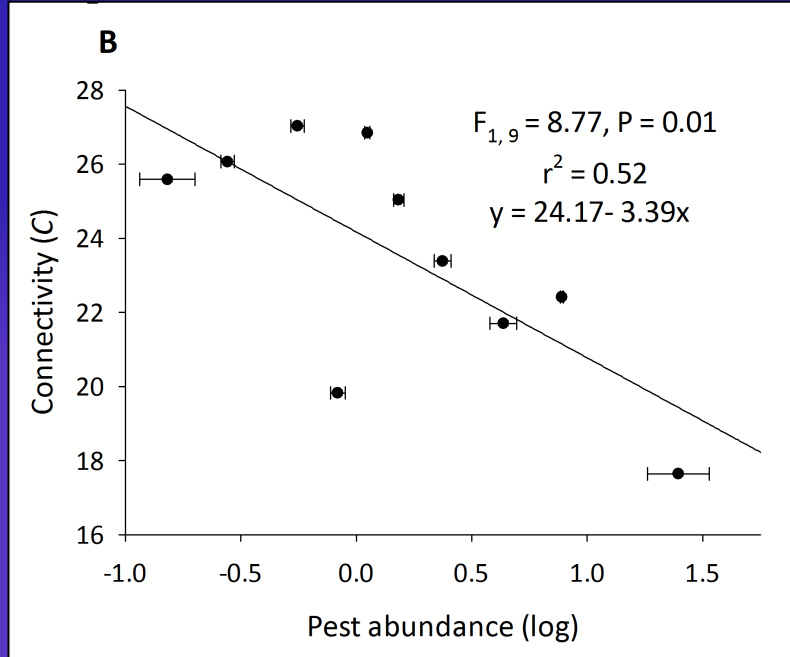
Function

Can we assess what species are at risk?

Community Network in Agroecosystems



Network Strength and Pest Populations



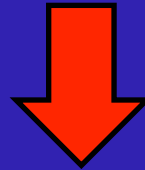
More linked
communities have fewer
pests



Using Community Ecology to Select Exposed Species



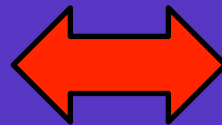
Bioinventory



Abundance and Incidence



Top corn consumers



Network connection to
corn feeders



Bioinventory of Corn Arthropods

Soil column, soil
surface, plant
foliage

18 farms across
eastern SD

Mid-vegetative
and anthesis

382 taxa collected
11,939 specimens

Lundgren et al. 2015. Spatial and numerical relationships of arthropod communities associated with key pests of maize. J Appl Entomol, in press
Welch and Lundgren. An exposure-based, ecologically driven framework for selection of indicator species for insecticide risk assessments. Food Webs, in review

Most Abundant Taxa

Taxa present in at least 50% of fields

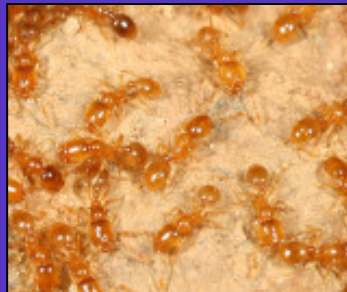
Herbivore

Rhopalosiphum padi
Frankliniella sp.



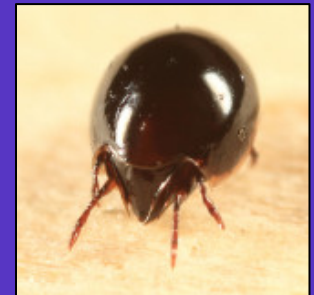
Predator

Solenopsis (Thief ant)



Detritivore

Oribatid mite



Corn Consumers

21 taxa were found to eat corn

Herbivore

Lygus lineolaris



Predator

Chrysoperla sp.



Detritivore

Lepidocyrtus sp. (Collembola: Entomobryidae)



Most Connected Species

Number of network connections to corn consumers

Herbivore

Frankliniella sp.
Malloewia sp. (Chloropidae)

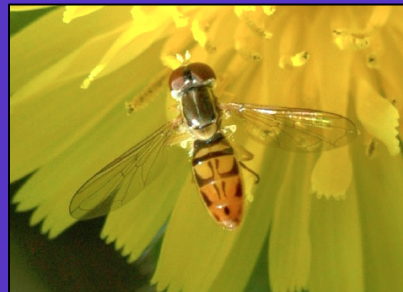


Predator

Chrysoperla sp.

Pollinator

Toxomerus sp. (Syrphidae)



Detritivore

Oribatid mite

Summary

A diverse community resides in corn, with numerous species that are trophically exposed to PIPs.

Based on abundance, corn consumption, and network connectivity, some ecologically relevant species that are:

Herbivore

Thrips
Lygus

Predator

Green lacewing
Ants
(*Orius* also scored highly)

Detritivore

Oribatid mite
Entomobryid Collembola

Bioinformatics as a Tool for RNAi Risk Assessment

Given the diversity of non-target species, can we

Reduce the number of species to test

Optimize hazard assessments to detect phenotypic changes

Unintended gene silencing in non-target organisms

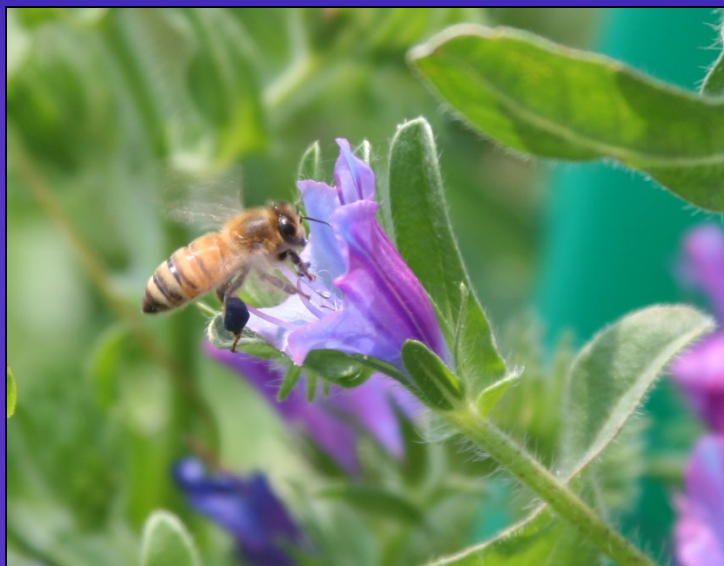
The Database

Toxins

101 pesticidal dsRNAs (and siRNAs)

57 gene targets

23 species of taxonomic targets

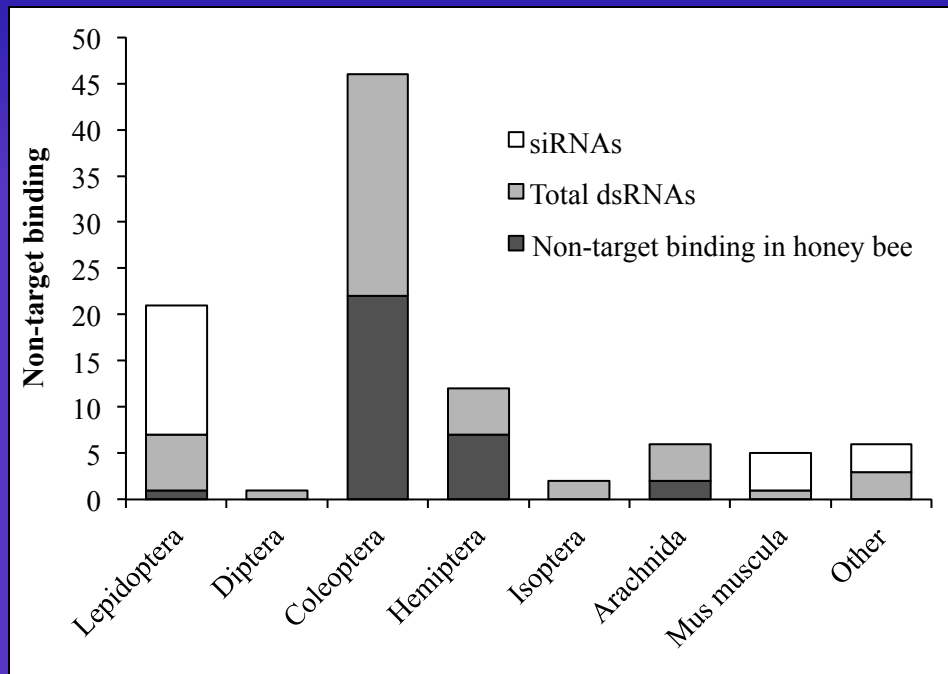


Searched for sequence
homologies (19/21, 20/21,
and 21/21 nt) in honey bee
genome

Off-target Gene Homologies

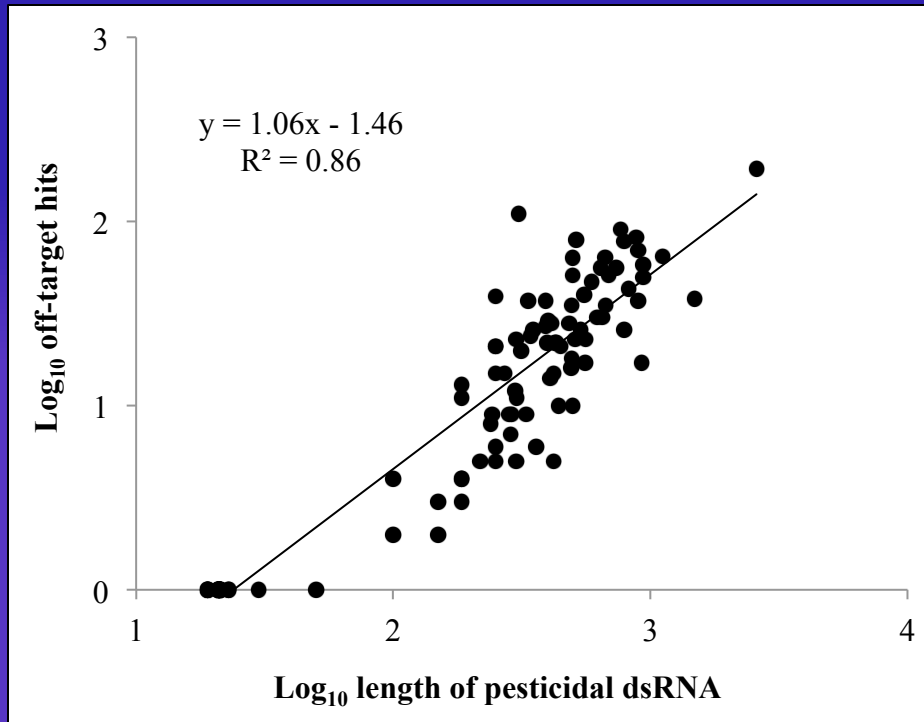
All of the the dsRNAs had at least one perfect sequence homology in honey bees

siRNAs had no gene homologies in bees



Taxonomy of target does not affect the likelihood of silencing the targeted gene in the honey bee.

Homeobox and developmental genes had a disproportionately high level of sequence homology with pesticidal dsRNAs



Longer dsRNAs
produced more off-
target hits

Conclusions

Pesticidal dsRNAs likely will find off-target binding sites in non-target organisms

In silico gene homologies does not imply phenotypic suppression

Predicted gene suppression can help to hone hazard assessments

Conclusions

Risks of pesticidal RNAi should be predictable and may be avoidable.

Filling the numerous knowledge gaps surrounding these risks will improve this predictability.

Now is the time to be asking these questions



Pests are a Symptom, NOT the Problem!

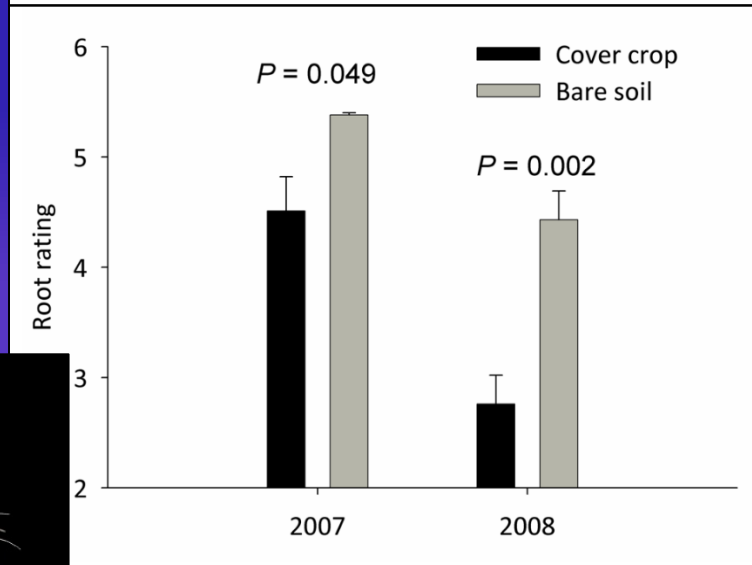
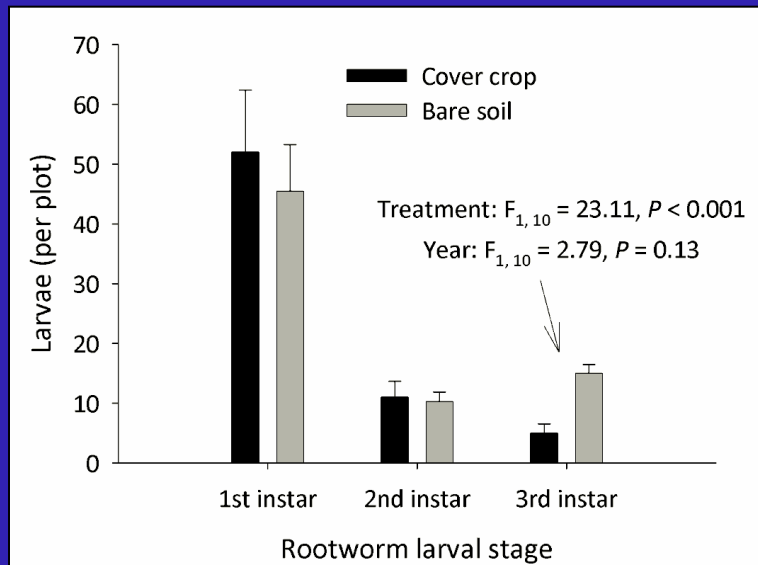
What causes pest problems?

High Disturbance Causes Pest Problems



Tillage

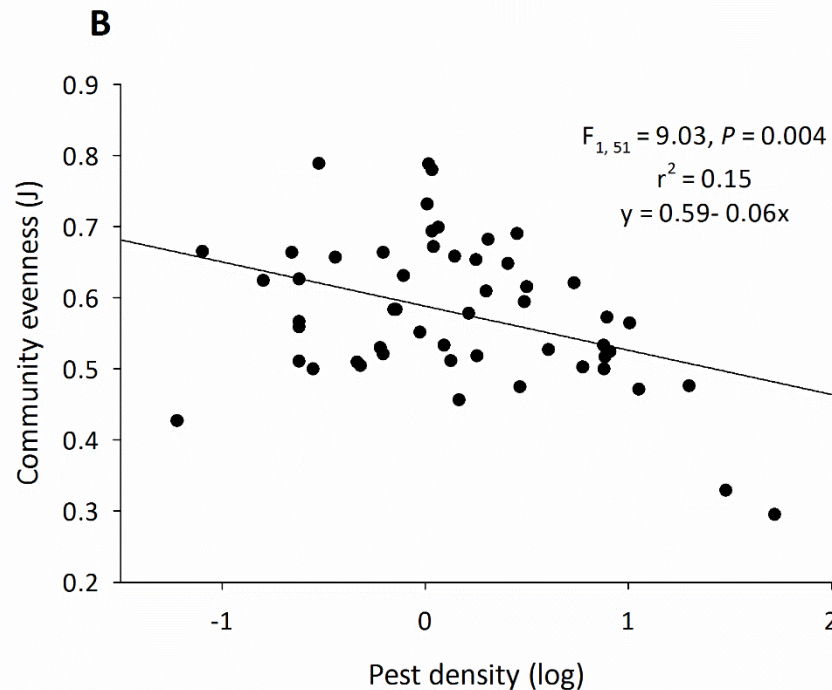
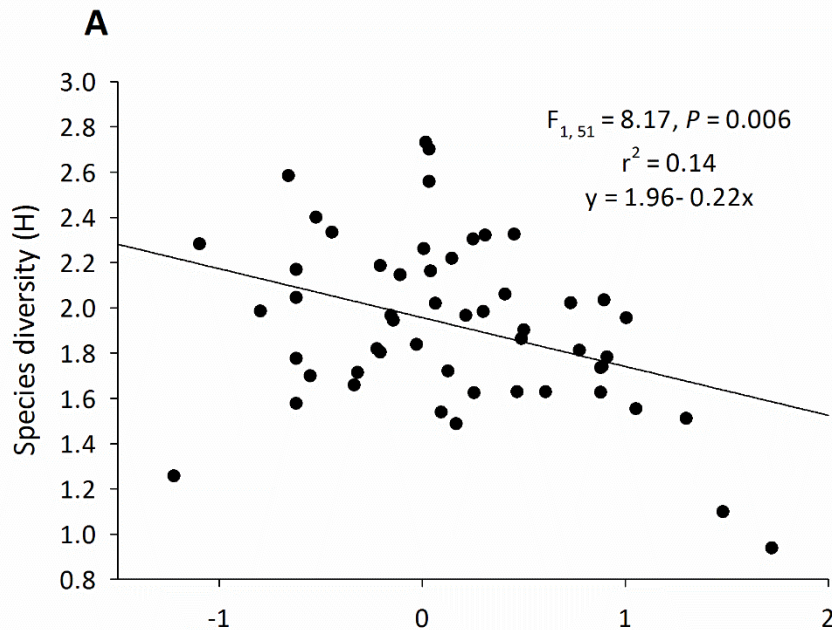
Brust et al. 1985. J Econ Entomol 78: 1389
Lehman et al. 2015. Sustainability 7: 988



Bare soil

Lundgren and Fergen 2010. Environ Entomol 39: 1816
Lundgren and Fergen 2011. Appl Soil Ecol 51: 9

Low Diversity Causes Pest Problems

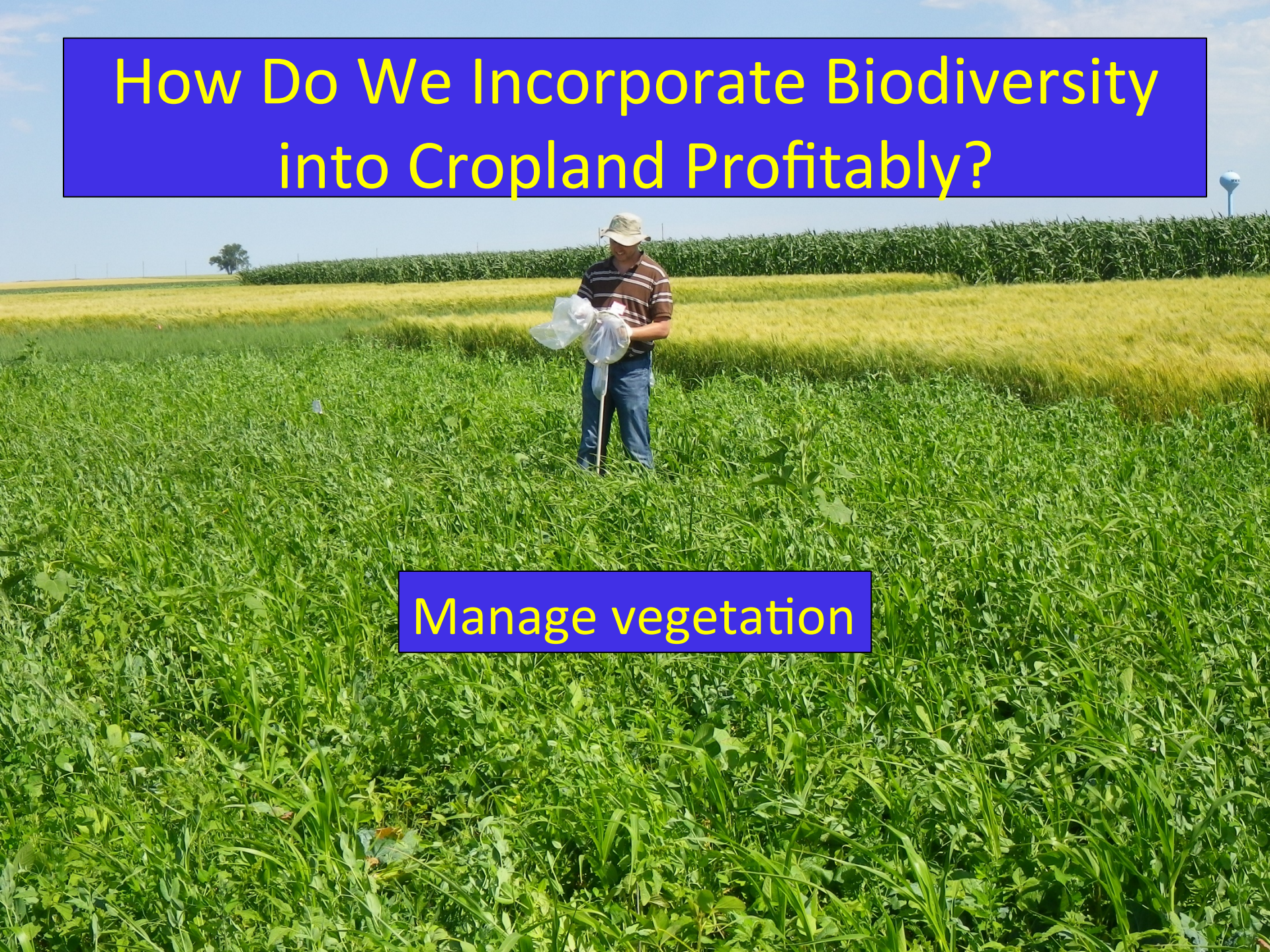


Not just species richness,
but the balance of species
within that community and
the interactions that they
have

Lundgren and Fausti. Trading biodiversity for pest problems.
Nature, in review

How Do We Incorporate Biodiversity into Cropland Profitably?

Manage vegetation



Increasing Diversity on Farms

Crop rotation

Intercropping

Smaller plots, more crops

Field margins

Cover crops

Weeds

Conservation strips

Andow 1991. Annu Rev Entomol 36: 561
Letourneau et al. 2011. Ecol Appl 21: 9

The Farmers are leading the way



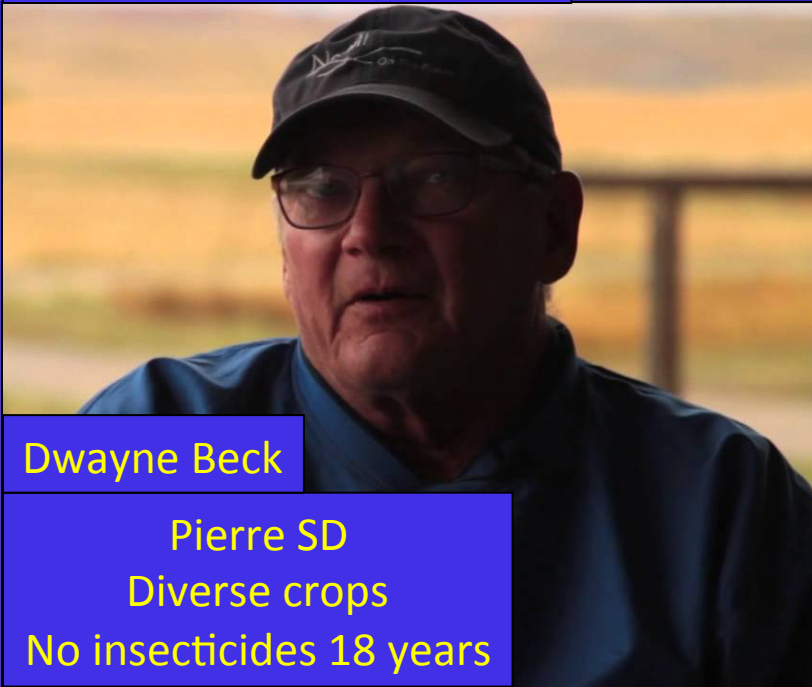
Gabe Brown

Bismarck ND
Diverse crops and livestock
No insecticides 28 years



Dave Brandt

Carroll, OH
Diverse crops
No insecticides 8 years



Dwayne Beck

Pierre SD
Diverse crops
No insecticides 18 years



Gail Fuller

Emporia KS
Diverse crops and livestock
No insecticides 8 years

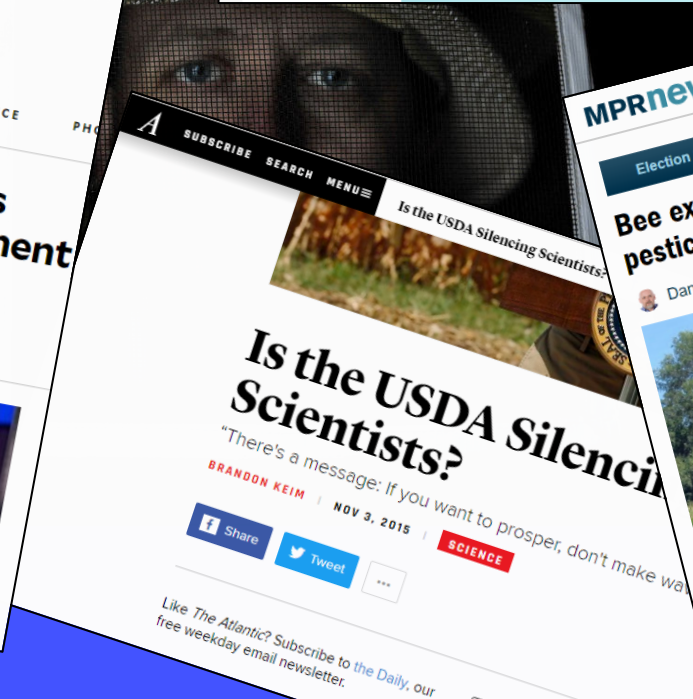


Translating Research to Practice

Research supports these ideas

It is diffuse
The endpoints are often wrong
There are numerous knowledge gaps

Why?



A New Way for Science to Help Bee Keepers and Farmers



www.ecdysis.bio



www.bluedasher.farm





Collaborators:
Scott Fausti
Jian Duan
Cable Hardin

Funding

USDA-NIFA
USDA-ARS



Technical help:
Janet Fergen
Greta Schen
Mike Bredeson
Chrissy Mogren
Ryan Bell
Claire Bestul
Kae Januschka
Chloe Kruse
Jacob Pecenka
Cally Strobel
Kelton Welch